

## Spatial Control of Crystal Nucleation in Agarose Gel

Carla Duffus, Philip J. Camp, and Andrew J. Alexander\*

*School of Chemistry, University of Edinburgh, The King's Buildings, West Mains Road, Edinburgh EH9 3JJ, U.K.*

Received June 25, 2009; E-mail: andrew.alexander@ed.ac.uk

Crystallization has long been a primary tool used by chemists for the production of solid samples, e.g., as a means of purification or for analysis by diffraction methods.<sup>1</sup> By control of various chemical and physical parameters such as temperature, solvent, or cosolutes, chemists have been able to engineer the growth of crystals of many substances.<sup>2,3</sup> Unfortunately, crystal growing can still be considered a black art since the initial stage of crystallization—nucleation—is a result of random fluctuations, and it is difficult to predict the effects of changing environment on the nucleation and subsequent growth.

Gels present many advantages as media for crystal growth: they are known to promote growth of larger, single crystals with fewer defects and different morphologies.<sup>4–6</sup> The suppression of convection currents and sedimentation in a gel produces an environment similar to microgravity, and so gels have become increasingly popular for growing crystals of proteins and other biological macromolecules.<sup>7</sup>

In this Communication, we demonstrate for the first time both spatial and temporal control of crystal nucleation in agarose gels using a recently discovered phenomenon of nonphotochemical laser-induced nucleation (NPLIN).<sup>8</sup> The method employs pulses of laser light at visible or near-infrared wavelengths and with relatively low powers to avoid photochemistry. The *peak* electric field of the light is sufficient to modify the free energy of prenucleating clusters, causing them to become supercritical and thereby subject to growth. The NPLIN effect was discovered by Garetz, Myerson, and co-workers<sup>8</sup> and has been demonstrated so far in aqueous or ethanol solutions for a range of substances such as urea,<sup>8</sup> glycine,<sup>9</sup> hen egg-white lysozyme,<sup>10</sup> and KCl.<sup>11</sup> We note that femtosecond laser light has also been used to induce nucleation, although the intensity of such light causes photochemical and photomechanical damage to solute and solvent.<sup>12</sup>

Supersaturated KCl–agarose gels were prepared by dissolving 0.12–0.75% w/w powdered agarose (Sigma-Aldrich, type I, A6013) in supersaturated (106%) aqueous solutions of KCl at 95 °C. The hot gel was then poured into vessels and allowed to cool to 23 °C and held at this temperature for about 30 min prior to shooting with a laser. A Nd<sup>3+</sup>:YAG laser was used, producing pulses of near-infrared light (1064 nm, 6 ns pulse width) in a 5.5 mm diameter beam. The power of the laser pulses was varied by passing the polarized light through a Glan-laser polarizer.

As a qualitative demonstration of NPLIN in gel, we have used a simple optical lithography technique to control the location of crystal nucleation. A thin (~2 mm) layer of gel (0.25% w/w) was prepared by pouring into a glass Petri dish. After cooling, a cutout mask was placed over the gel which was then subjected to a series of laser pulses that were raster scanned across the area of the mask. The results are shown in Figure 1, which shows that crystals are only observed where the light could pass through the mask. It is well-known that mechanical shock can cause nucleation in supersaturated solutions.<sup>1</sup> By repeat experiments, we verified (to within

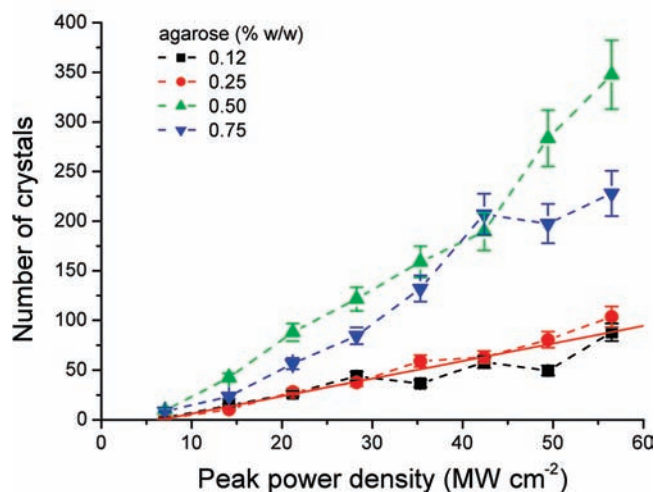
~100 μm) that no nucleation occurs beyond the edge of the mask, e.g., due to acoustic shockwaves.



**Figure 1.** Photograph showing spatial control of laser-induced nucleation of KCl in an agarose gel. The pattern, showing the word “LASER” with stars above and below, was obtained using an optical mask. No crystals were observed in the regions that were masked. The diameter of the dish was 9 cm (see text for further details).

The control we have demonstrated is limited by localization of the laser pulse. The nucleation within the illuminated region is stochastic, however, since it depends on the distribution of prenucleating (subcritical) clusters. We have observed that simple focusing of the beam can lead to damage of the gel at higher laser powers. We have also observed that two low-power beams can be combined to induce nucleation where they are crossed, opening the route to three-dimensional control of nucleation. The simple technique could be easily developed to a wide range of systems, such as nonaqueous gels or in droplets.<sup>5,13</sup> Recent developments in optical methods could be applied to improve localization within a solid matrix.<sup>14</sup>

To measure the dependence of nucleation on laser pulse power, aliquots of gel were dispensed into small vials (~3 cm<sup>3</sup>), each of which was shot with a single laser pulse. After approximately 10 min, the vials were photographed and crystals counted. The results (Figure 2) show an apparent threshold power (~7 MW cm<sup>-2</sup>) below which no crystals were nucleated, in agreement with previous results in solution.<sup>11</sup> At the present time, there is no clear explanation for a power threshold in solution or in gel, and further experiments are underway to investigate fully this phenomenon. At low agarose concentrations (≤0.25% w/w), the number of crystals nucleated increases approximately linearly with peak laser power. A similar dependence of fractions of aqueous samples nucleated versus laser power was reported; in those experiments, a single crystal per vial



**Figure 2.** Plot showing the number of crystals counted resulting from a single laser pulse as a function of the peak power of the pulse.

was produced, on average.<sup>11</sup> The relatively higher number of crystals nucleated in agarose can be explained in an approximate theory by an effective lowering of the energetic barrier to nucleation due to the presence of agarose. To this end, we model the crystal nuclei as homogeneous spherical domains immersed in a continuum solvent. For *homogeneous* nucleation of a subcritical cluster of radius  $r$ , the classical free energy of formation can be written

$$\Delta G_{\text{hom}}(r, E) = 4\pi r^2 \gamma - \frac{4}{3}\pi r^3 (A \ln S + aE^2) \quad (1)$$

where  $E$  is the electric field due to the laser pulse;  $\gamma$  is the cluster interfacial tension;  $S$  is supersaturation ( $S = C/C_{\text{sat}} = 1.06$ ); and  $a = 1.7832 \times 10^{-12} \text{ F M}^{-1}$  depends on the dielectric permittivities of KCl and water (at 1064 nm,  $\epsilon_{\text{KCl}} = 2.1897$  and  $\epsilon_{\text{water}} = 1.7535$ ).<sup>11,15</sup> The term  $aE^2$  arises from interactions between the electric field and the electronic polarizability of the cluster. The parameter  $A = \rho RT/M = 6.553 \times 10^7 \text{ J m}^{-3}$ , where  $\rho$  is the mass density and  $M$  is the molar mass of KCl and  $R$  and  $T$  are the molar gas constant and temperature, respectively. For *heterogeneous* nucleation on a planar surface, eq 1 becomes

$$\Delta G_{\text{het}}(r, E) = f(\theta) \Delta G_{\text{hom}}(r, E) \quad (2)$$

where  $f(\theta) = (2 + \cos \theta)(1 - \cos \theta)^2/4$ , and  $\theta$  is the contact angle between cluster and substrate.<sup>1</sup> The barrier to nucleation can only be reduced by the crystal–agarose interaction; i.e.,  $f(\theta) \leq 1$ , resulting in more nucleating clusters. The same holds true for other surface geometries such as pores or troughs.

From previous experiments of NPLIN on aqueous KCl solutions, we have determined  $\gamma = 2.19 \text{ mJ m}^{-2}$ .<sup>11</sup> Taking averages over the classical Boltzmann distribution,  $\exp(-\Delta G_{\text{het}}/k_B T)$ , we calculated numbers of clusters that become supercritical as a function of laser power. The model predicts a linear dependence of numbers of nuclei versus peak power density. However, the calculated numbers of crystals are orders of magnitude too high even for the case of complete dewetting ( $\theta = 180^\circ$ ,  $f(\theta) = 1$ ; equivalent to homogeneous nucleation). This result suggests that our model underestimates  $\gamma$  or overestimates  $S$ , or both. In the absence of independent estimates of  $\theta$ , we assume homogeneous nucleation ( $\theta = 180^\circ$ ) and explore possible effective parameters,  $\gamma^{\text{eff}}$  and  $S^{\text{eff}}$ , that fit the data at 0.25% w/w agarose content. Assuming  $S = 1.06$ , we find that  $\gamma^{\text{eff}} = 5.13$

$\text{mJ m}^{-2}$  gives an excellent straight line fit. Alternatively, fixing  $\gamma = 2.19 \text{ mJ m}^{-2}$ , the data can be fitted with  $S^{\text{eff}} = 1.016$ . These two different fits are indistinguishable and are shown as the straight line in Figure 2. Note that the calculations have been shifted along the power axis by  $7 \text{ MW cm}^{-2}$  to reproduce the as-yet unexplained experimental threshold.

There is no obvious explanation for why  $\gamma^{\text{eff}}$  would be greater than  $\gamma$  in solution. A reduction in the effective supersaturation, defined by  $S^{\text{eff}} = C^{\text{eff}}/C_{\text{sat}}^{\text{eff}}$ , could arise from one or both of the following effects. (i) The saturation concentration of KCl in gel may be higher than that in pure water ( $C_{\text{sat}}^{\text{eff}} > C_{\text{sat}}$ ), due to a stabilization of solvated ions through long-range Coulombic interactions with charged groups (e.g., sulfate<sup>16</sup>) on the agarose-gel surface. (ii) The concentration of ions in solution and available for nucleation may be reduced ( $C^{\text{eff}} < C$ ), due to a sequestering of ions within the gel matrix.<sup>17</sup>

At higher laser powers, nonlinearity in the numbers of crystals nucleated may indicate the onset of different mechanisms. We also note that the 0.75% w/w agarose produced fewer crystals than 0.5% w/w agarose. This may be attributed to changes in the gel structure at higher agarose concentrations, including changes in pore size and connectivity.<sup>18</sup>

In summary, we have demonstrated remarkable temporal control (on a nanosecond time scale) and spatial control (to within  $\sim 100 \mu\text{m}$ ) of crystal nucleation. Results can be fitted using an approximate continuum theory, but further work is required to understand the atomic-scale mechanism for NPLIN. We are now investigating NPLIN in other solvent and gel systems. The new method described here shows true potential for use as a routine tool in laboratory growth of crystals, e.g., for controlled growth of single crystals of proteins or other materials for structure analysis.

**Acknowledgment.** A.J.A. thanks The Royal Society and the EPSRC (EP/G067546/1) for funding.

## References

- (1) Mullin, J. W. *Crystallization*, 4th ed.; Butterworth-Heinemann: Oxford, 2001.
- (2) Lee, A. Y.; Lee, I. S.; Dette, S. S.; Boerner, J.; Myerson, A. S. *J. Am. Chem. Soc.* **2005**, *127*, 14982–14983.
- (3) Cheng, C. M.; LeDuc, P. R. *J. Am. Chem. Soc.* **2006**, *128*, 12080–12081.
- (4) Henisch, H. K. *Crystals in Gels and Liesegang Rings*; Cambridge University Press: New York, 1988.
- (5) Doxsee, K. M.; Chang, R. C.; Chen, E.; Myerson, A. S.; Huang, D. *J. Am. Chem. Soc.* **1998**, *120*, 585–586.
- (6) Li, H.; Estroff, L. A. *J. Am. Chem. Soc.* **2007**, *129*, 5480–5483.
- (7) Biertümpfel, C.; Basquin, J.; Suck, D.; Sauter, C. *Acta Crystallogr.* **2002**, *D58*, 1657–1659.
- (8) Garetz, B. A.; Aber, J. E.; Goddard, N. L.; Young, R. G.; Myerson, A. S. *Phys. Rev. Lett.* **1996**, *77*, 3475–3476.
- (9) (a) Zaccaro, J.; Matic, J.; Myerson, A. S.; Garetz, B. A. *Cryst. Growth Des.* **2001**, *1*, 5–8. (b) Garetz, B. A.; Matic, J.; Myerson, A. S. *Phys. Rev. Lett.* **2002**, *89*, 175501.
- (10) Lee, I. S.; Evans, J. M. B.; Erdemir, D.; Lee, A. Y.; Garetz, B. A.; Myerson, A. S. *Cryst. Growth Des.* **2008**, *8*, 4255–4261.
- (11) Alexander, A. J.; Camp, P. J. *Cryst. Growth Des.* **2009**, *9*, 958–963.
- (12) (a) Yoshikawa, H. Y.; Hosokawa, Y.; Masuhara, H. *Jpn. J. Appl. Phys.* **2006**, *45*, L23–L26. (b) Nakamura, K.; Hosokawa, Y.; Masuhara, H. *Cryst. Growth Des.* **2007**, *7*, 885–889.
- (13) Draper, N. D.; Bakhoun, S. F.; Haddrell, A. E.; Agnes, G. R. *J. Am. Chem. Soc.* **2007**, *129*, 11364–11367.
- (14) (a) Campbell, M.; Sharp, D. N.; Harrison, M. T.; Denning, R. G.; Turberfield, A. J. *Nature* **2000**, *404*, 53–56. (b) Andrew, T. L.; Tsai, H. Y.; Menon, R. *Science* **2009**, *324*, 917–921.
- (15) Lide, D. R., Ed. *Handbook of Chemistry and Physics*, 88th ed.; CRC Press: Boca Raton, FL, 2007.
- (16) Podesva, J.; Prochazka, O.; Medin, A. *Polymer* **1995**, *36*, 4967–4970.
- (17) Valtchev, V. P.; Bozhilov, K. N. *J. Am. Chem. Soc.* **2005**, *127*, 16171–16177.
- (18) Maaloum, M.; Pernodet, N.; Tinland, B. *Electrophoresis* **1998**, *19*, 1606–1610.

JA905232M